

Signaling pathways involved in chronic myeloid leukemia pathogenesis: The importance of targeting Musashi2-Numb signaling to eradicate leukemia stem cells

Foruzan Moradi¹, Sadeqh Babashah^{1*}, Majid Sadeghizadeh¹, Arsalan Jalili^{2,3}, Abbas Hajifathali^{2,4}, Elham Roshandel^{4,5}

¹ Department of Molecular Genetics, Faculty of Biological Sciences, Tarbiat Modares University, Tehran, Iran

² Hematopoietic Stem Cell Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran

³ Department of Stem Cells and Developmental Biology at Cell Science Research Center, Royan Institute for Stem Cell Biology and Technology, ACECR, Tehran, Iran

⁴ Taleghani Hospital, Shahid Beheshti University of Medical Sciences, Tehran, Iran

⁵ Department of Hematology, School of Medical Sciences, Tarbiat Modares University, Tehran, Iran

ARTICLE INFO

Article type:

Review article

Article history:

Received: May 15, 2018

Accepted: Nov 15, 2018

Keywords:

BCR-ABL1

Chronic myeloid leukemia

Cancer stem cells

Signaling pathways

Self-renewal

Targeted therapy

ABSTRACT

Objective(s): Chronic myeloid leukemia (CML) is a myeloid clonal proliferation disease defining by the presence of the Philadelphia chromosome that shows the movement of BCR-ABL1. In this study, the critical role of the Musashi2-Numb axis in determining cell fate and relationship of the axis to important signaling pathways such as Hedgehog and Notch that are essential for self-renewal pathways in CML stem cells will be reviewed meticulously.

Materials and Methods: In this review, a PubMed search using the keywords of Leukemia, signaling pathways, Musashi2-Numb was performed, and then we summarized different research works.

Results: Although tyrosine kinase inhibitors such as Imatinib significantly kill and remove the cell with BCR-ABL1 translocation, they are unable to target BCR-ABL1 leukemia stem cells. The main problem is stem cells resistance to Imatinib therapy. Therefore, the identification and control of downstream molecules/ signaling route of the BCR-ABL1 that are involved in the survival and self-renewal of leukemia stem cells can be an effective treatment strategy to eliminate leukemia stem cells, which supposed to be cured by Musashi2-Numb signaling pathway.

Conclusion: The control of molecules /pathways downstream of the BCR-ABL1 and targeting Musashi2-Numb can be an effective therapeutic strategy for treatment of chronic leukemia stem cells. While Musashi2 is a poor prognostic marker in leukemia, in treatment and strategy, it has significant diagnostic value.

Please cite this article as:

Moradi F, Babashah S, Sadeghizadeh M, Jalili A, Hajifathali A, Roshandel E. Signaling pathways involved in chronic myeloid leukemia pathogenesis: The importance of targeting Musashi2-Numb signaling to eradicate leukemia stem cells. Iran J Basic Med Sci 2019; 22:581-589. doi: 10.22038/ijbms.2019.31879.7666

Introduction

Cancer refers to a group of diseases, which irregular cell growth is their outstanding feature; it can predominantly invade and spread cells from the primary site to other parts of the body (1, 2). When immature blood cells in the bone marrow (progenitor cell or precursor) grow uncontrollably and prevent the production of healthy blood cells, a type of cancer called leukemia is created, which is the unregulated proliferation of cells and disrupting of the bone marrow functions leading to an untimely death if left untreated (3, 4). Leukemia can be acute or chronic; each of them is divided into lymphoid and myeloid lineages according to their origins (5, 6). Chronic myeloid leukemia (CML) is a monoclonal proliferating malignant associated with myeloid lineage (7). CML is diagnosed by detecting increased clonal hematopoietic stem cell(s) (HSCs) that has a shift between chromosomes 9 and 22. The resulting chromosome is called Philadelphia (Ph). The inverted Abelson murine leukemia viral oncogene homolog 1 (ABL1) gene located in 9p34 is placed in breakpoint

cluster region protein (BCR) gene located in 22q11 zone and results in the formation of BCR-ABL1 fusion gene that creates tyrosine kinase BCR-ABL1, which is always active and is the activator of several molecular pathways. Ultimately, this gene leads to abnormal cell adhesion, increased proliferation, and inhibition of apoptosis (8-10). CML clinical courses are divided into chronic phase, accelerated phase and blastic phase (8, 11). It develops slowly, while gradually indicates an increase in blood and bone marrow blast percentage, and the accelerated phase will evolve as a blast crisis phase (12, 13), which progressive resistance to therapy happens during this stage (14, 15).

Tyrosine kinase that is an important mediator of the signaling cascade plays a key role in diverse biological processes like growth, differentiation, metabolism and apoptosis. For decades, it had been postulated that Imatinib mesylate, an inhibitor of the specific BCR-ABL1 tyrosine kinase, is the most effective and selective therapeutic strategy, which had been developed as the first molecularly targeted therapy (16-19). Imatinib

*Corresponding author: Sadeqh Babashah. Department of Molecular Genetics, Faculty of Biological Sciences, Tarbiat Modares University, Tehran, Iran. Tel: +98-21-82884468; Fax: +98-21-82884717; Email: babashah@modares.ac.ir

typically targets abnormal cells in leukemia, but they are not effective on leukemia stem cells (LSCs) (20). So, these cells remain intact during the conventional cancer therapies, such as chemotherapy and radiation therapy, leading to tumor recurrence and metastasis. Therefore, recognizing the pathways of proliferation, self-renewal, and survival of normal and malignant stem cells can lead to a better understanding of cancer and to find new therapies, especially by targeting cancer stem cells (21).

Accumulating evidence has demonstrated that all of the important signaling pathways related to survival of LSCs can be activated by oncogene BCR-ABL1; therefore, this oncogene creates self-renewal property in LSCs (22, 23). However, BCR-ABL1 oncogene alone cannot be responsible for the self-renewal ability of the committed progenitors for transforming them and will use self-renewal properties of the cells such as HSCs (22, 24, 25). The signaling pathways like Hedgehog (Hh) (26, 27), Wnt/ β -catenin (28), Notch (25), Alox5 (30, 31), Foxo (65-67), and Ras (62) may play roles in differentiation and survival of LSCs (35, 36). Beside them, Musashi2 (Msi2)-Numb signaling axis is a molecular pathway, which is correlated with other signaling pathways such as Hh and Notch involved in regulating the self-renewal properties of LSCs (37-39). The Msi2 elimination can increase Numb expression levels. This increment of Numb through the key genes of Hh and Notch signaling pathways can decrease the number of LSCs. Hence, the importance of targeting the Msi2-Numb signaling axis to eradicate LSCs will be discussed in details. Regulating these signaling pathways in cancer cells is disrupted; therefore, study in this area may offer some options in the treatment of leukemia.

Materials and Methods

The following data was collected by electronic databases, including PubMed, Scopus, ScienceDirect and Cochrane library, Google Scholar, and Web of Science. All types of relevant studies, original articles, books and abstracts were included and their results were reviewed and reported until 2018.

Results

The resistance of CML stem cells to BCR-ABL1 kinase inhibitor called Imatinib

The development of Imatinib mesylate, as a tyrosine kinase inhibitor, that is used as a first-line treatment in newly diagnosed CML patients had brought significant successes in the treatment of patients with CML (40-42). Using Imatinib 400 mg/day for patients with chronic disease, led to 70% cytogenetic improvement and 3-year survival. This level is lower in patients with accelerated phase or blast crisis (43, 44). Although continuous Imatinib therapy, especially in CML stage, leads to permanent responses, about 20 to 30 percent of CML patients show resistance to Imatinib therapy and only a few percent will improve and that is because of the secondary mutations in BCR-ABL1 and generating kinases. Patients are resistant to the Imatinib therapy and only a few percent will improve (45). Hence, there is no evidence to show that patients receiving Imatinib can safely stop using it since most of the patients who had interrupted their treatments, have faced with early recurrences of the molecular and cytogenetic level even after remission (43, 44, 46) and therefore these patients

have to endure a lifelong therapy (47-50). Conventional treatments such as chemotherapy and radiation therapy that target dividing cells and decrease tumor mass cannot prevent tumor regrowth because these approaches are not capable of killing the cancer stem cells (21). Another problem in conventional treatment procedures is the low specifications, which makes the drugs kill the natural cells beside the tumor cells, and this is one reason for the ineffectiveness and side effects of these methods (51, 52). This is supported since BCR-ABL-positive LSCs remain intact even after long-term imatinib therapy and can cause relapse of the disease. These findings suggest that inhibition of BCR-ABL tyrosine kinase activity alone is insufficient to eradicate LSCs (53-58). Therefore, development of efficient therapeutic strategies capable of targeting key genes involved in self-renewal signaling pathways in LSCs would provide noticeably improved therapeutic benefits to patients suffering CML (59-63) (Figure 1). It was clear that self-renewal property of stem cells can create drug-resistant and recurrence of disease in different cancers (64, 65). It is believed that genetic changes cause deregulation of stem cells that in turn will provide unlimited self-renewal potential for them. As a result, aberrant self-renewal of stem cells provides a prerequisite for initiation, progression, and resistance to cancer (66, 67). Since HSCs and CML stem cells are capable of self-renewal, it is not inconceivable that several signaling pathways that are involved in the regulation of standard stem cells have also roles in cancer stem cell biology, and in this context, there is evidence showing that most of the pathways that are classically associated with cancer may participate in the regulation of normal stem cell development (36, 58, 68, 69).

Musashi2-Numb signaling axis

Musashi family involves a group of RNA-binding proteins that are expressed in stem cells and invasive tumors (70). In mammalian, two isoforms of this family called Msi1, and Msi2 have been identified (37). Msi1 gene is located on chromosome locus 12q24 and has 22 exons, and the Msi2 gene has 19 exons and is placed on chromosome locus 17q22. Both isoforms are identified by RNA motifs (RRM), which have high homology in their sequences (72). Experimental investigations show special specification in the binding region of both proteins (71). The similarity in the amino acid surface at the binding region to RNA is 81% for the first RRM and 93% for the second RRM, while 100% similarity is reported for the region of binding, which includes octapeptide conserved sequences (RGFGVTF) (72). Musashi proteins affect asymmetric cell division, stem cell functions and identifying cell fate in different somatic cells (73, 74). However, their expression patterns in diverse tissues are different. Msi1 in neural stem cells and periventricular progenitor cells in the embryo and also after birth are rich. It is also active in the mammalian brain, gastrointestinal tract, mammary gland, skin, and tumorigenesis (75-82). Overexpression of Msi1 in the advanced stages of the disease and also the prognosis associated with colon and breast cancers (83, 84), urothelial, esophageal, cervical cancer as well as central nervous system tumors, including glioma, medulloblastoma, and ependymoma is recorded (85-90). Some tissues like central nervous system, and nervous stem cells (NSCs) show the simultaneous activity of Msi1 and Msi2 (76). What is important is

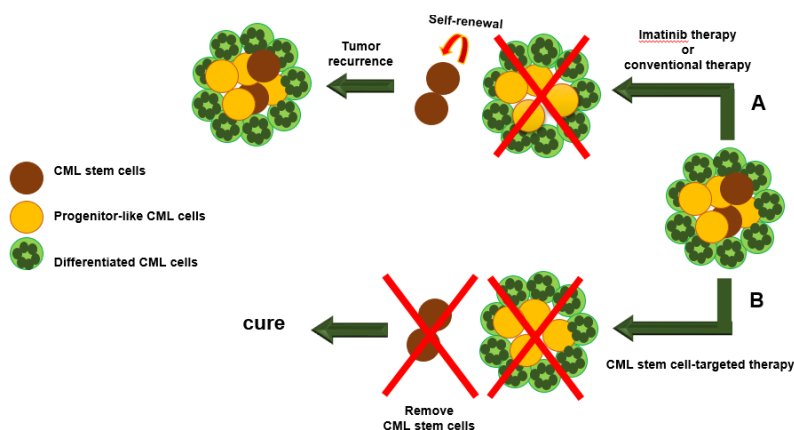


Figure 1. Chronic myeloid leukemia (CML) stem cells are resistant to conventional treatments or breakpoint cluster region protein- Abelson murine leukemia viral oncogene homolog 1 (BCR-ABL) kinase inhibitor called Imatinib. (A) Conventional treatments such as chemotherapy and radiation therapy that target dividing cells and decrease tumor mass cannot prevent tumor regrowth because these approaches are not capable of killing the cancer stem cells. Although Imatinib can inhibit an integrated protein called BCR-ABL1 and decrease the number of CML cells, this inhibitor cannot target the CML stem cells, and this leads to recurrence of the disease. (B) If there would be a method that specifically targets CML stem cells and leads to decrease the self-renewal of CML stem cells, the remaining cells would not be able to support their cancer identity. This approach may avoid drug resistance and recurrence of disease in the patients with CML who are treated with Imatinib. This figure was adapted from reference

the point that Msi1 expression in HSCs and progenitor cells are very small and negligible when compared to Msi2 (38, 70), while Msi2 in hematopoietic systems has striking expression (73), and it is recognized as a primary regulator of the HSCs (70, 91) and LSCs (38, 70). Several studies have also reported the relationship of the Msi2 family members to solid tumor pathogenesis (85, 92, 93).

Msi2 gene has been identified as a significant indicator for myeloid leukemia (94). Not only the overexpression of Msi2 is a weak marker in the progression of human CML (38), but it is also related to the rapid progression and poor prognosis in myeloid leukemia (38, 70, 95). Previous studies have shown that Msi2-Numb developmental pathway plays a critical role in the retention of CML stem cell functions and the activities of such pathways would lead to proliferation, development, and survival of LSCs. In other word, Msi2 gene plays a significant role in complex regulation pathways that are involved in self-renewal, and proliferation of HSCs such as Ras, mitogen-activated protein kinase (MAPK), Cycline D1, Myc (70), homeobox genes Meis1, HOXA9, HoX10, Hh and Notch pathways and possibly the other pathways as well (91). Msi2 can control the translation by specific sequence interactions with 3'UTR of target mRNAs in the different stem cells (96). In this regard, studies of Ito *et al.* and Kharas *et al.* in 2010 were the first studies that have shown the relationship between the Msi2 inducer of the Msi2-Numb pathway and hematopoietic malignancy, and their investigations indicated the Msi2 portion and its regulative interactions with leukemia in a stable and definitive manner (38, 70). The findings of these two reports are a primary step in treating aggressive leukemia. Using *in vivo* CML models in chronic phase and blast crisis (13) suggested that Msi2-Numb can be a novel target for leukemia treatment since it can control CML stem cells differentiation and apoptosis (38). In another study, Zhang *et al.* demonstrated that Msi2 knockdown inhibited leukemic cell proliferation and promoted cell apoptosis involving the MAPK signaling pathway. Their study provided novel insight into the

mechanisms of leukemogenesis (73). They investigated Msi2 expression at protein levels in K562, KG-1a, HL-60, THP-1, OCI-AML3 and U937 cell lines. According to their study, western blotting analyses showed high expression of Msi2 protein in KG-1a and K562 cells, and low expression in U937 and OCI-AML3 cells (Table 1). Kharas *et al.* showed high Msi2 expression levels in 6 cell lines associated with acute myeloid leukemia (AML) called Nomo-1, Skm-1, U937, NB4, Mono Mac 6, THP1 and low Msi2 expression levels in OCI-AML3 cell line (Table 2). The data are provided in the Tables according to references (70, 73).

Kharas *et al.* analyzed the gene expression of 436 patients with AML and reported that Msi2 expression level (as an independent prognosis marker) is related directly to decreased survival (70). They also reported deregulation of the key genes that control the self-renewal and cell fate in HSCs and may play a critical role in leukemia progression, and one of these adverse regulations is associated with Msi2-Numb signaling axis (70). This axis is significantly involved in regulating the cell self-renewal properties (37-39). They tested Msi2 expression inducer called doxycycline both in *in vitro* and *in vivo*; they observed that Msi2 expression in laboratory is at the highest level during the time of

Table 1. Msi2 expression in human leukemic cell lines. According to the results of Zhang and coworkers [73] Western blotting analyses showed high expression of Msi2 protein in KG-1a and K562 cells, and low expression in U937 and OCI-AML3 cells

Leukemia type	Cell line	Msi2 relative expression
CML	K562	Upregulated
AML	KG-1a	Upregulated
AML	HL-60	Upregulated
AML	THP-1	Upregulated
AML	OCI-AML3	Downregulated
AML	U937	Downregulated

Table 2. Msi2 expression in human leukemic cell lines. According to the results of Kharas and colleagues [70] Western blotting analyses showed the expression levels of Msi2 protein in 7 different leukemic cell lines

Leukemia type	Cell line	Msi2 relative expression
AML	Nomo-1	Upregulated
AML	Skm-1	Upregulated
AML	U937	Upregulated
AML	NB4	Upregulated
AML	Mono Mac 6	Upregulated
AML	THP1	Upregulated
AML	OCI-AML3	Downregulated

forming the hematopoietic colonies with immature myeloid phenotype, and Msi2 expression *in vivo* caused expansion of HSCs and short-term progenitor cells. They also conjugated Msi2 inducer doxycycline with BCR-ABL1 oncogenes and injected to mice. Consistent with other reports, they observed that Msi2 in immature myeloid leukemia in which blast crisis had been reconstructed was induced. Following these studies, expression of this gene in human samples with CML was also investigated (33 patients in blast crisis phase and 57 cases in chronic phase) and the results showed that Msi2 expression at progressive CML stages (blast crisis) is placed on the higher levels to primary levels of CML (chronic phase) and this overexpression of Msi2 has an inverse relationship with the Numb gene in the both blast crisis and chronic phase. Ito and coworkers (38) also indicated that there is a relationship between overexpression of Numb and decreasing of the leukemia cells in mouse models; they suggested that Numb cannot disperse the disease considerably. These results indicated that Numb

levels might prevent the progression of CML and induce differentiation in leukemia stem cells. Besides inducing the Numb expression, the results showed that Msi2 gene inhibition by shRNA1 would significantly decrease *in vivo* leukemia growth and retention rate, especially in blast phase. Similar to Numb inducing, Msi2 inhibition can also induce differentiation in leukemia cells and inhibit the ability of proliferation and distribution. A possible mechanism that is important in this area is that the inhibition effects of Msi2 elimination of LSCs may be related to Numb regulative effects, which are a determinant factor in cell fate. The Msi2 elimination can increase Numb expression levels and Numb may remove CML stem cells. These findings are possibly related to other signaling pathways that finally will decrease the numbers of LSCs.

Msi2 can inhibit the translation by binding to 3'UTR of Numb mRNA. By inhibiting the Msi2, the Numb gene will increase, and this increment of Numb is related to other signaling pathways like Hh and Notch. Finally, it will decrease the number of LSCs. By ubiquitination and destruction with the proteasome, Numb can inhibit Notch intracellular domain (NICD) and Gli that are respectively the components of Notch and Hh pathways. It is worth noting that Hh and Notch pathways are two important pathways that play significant roles in the self-renewal of resistant cancer stem cells. Therefore, by inhibiting these molecular ways, it is expected that cancer cell growth will be decreased and LSCs can be directed to apoptosis (38, 70, 74, 95) (Figure 2 A). Ito *et al.* in the mouse models that were in blastic phase indicated that Numb dramatically expressed at lower levels, while Msi2 expression in these models was particularly high (70). Kharas *et al.* presented the Numb gene expression at the protein levels (with Immunoblot) in LAMA-84 cell line (which is a cell line derived from blastic phase of CML patients) after transfection of

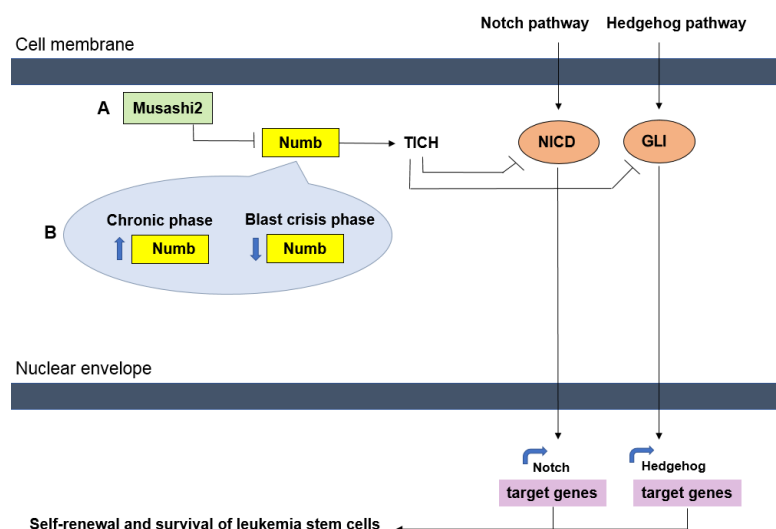


Figure 2. The Musashi2 (Msi2) gene is a major factor in normal hematopoietic and leukemia stem cells. (A) The elevated level of Msi2 leads to the downregulation of the cell-fate determinant, Numb gene, which probably relates to other signaling pathways like Hedgehog and Notch. Numb may decrease the number of Leukemia stem cells via ITCH factor: ITCH ubiquitinates Notch intracellular domain (NICD) and Gli that are respectively the component of Notch and Hedgehog pathways. It is worth noting that Hedgehog and Notch pathways are important pathways that play significant roles in the self-renewal of resistant cancer stem cells. (B) The continued suppression of Numb gene is required to maintain the blast crisis in CML. While the increasing of Numb expression or the inhibition of Msi2 expression might lead to inhibition of the Notch signaling pathway. Therefore, the CML blastic phase is suppressed. This figure was drawn by the writer to summarize the pathways involved in cell fate

siRNA against Msi2 (70). In their study, the inverse relationship between Numb and Msi2 genes was clearly evident. These reports indicate an inverse association of Msi2 gene and gene expression levels of Numb.

Hedgehog signaling pathway

Hh signaling pathway was first identified in *Drosophila* for patterning the early embryo. Several studies show that Hh signaling pathways may regulate the cell fate and maintain the stem cell/progenitor cells (97, 98). This highly conserved developmental pathway is responsible for regulating the normal HSCs and CML stem cells (98-101). Hh pathway is active in the embryonic hematopoiesis, but its level will decrease after birth; nevertheless, this channel will be increased again in patients with CML. This pathway is recognized as a functional pathway in the LSCs and any disruption in this way would prevent CML progression. The importance of Hh signaling pathway in carcinogenesis is attributed to self-renewal of stem cells. Hh pathway is activated by binding the protein ligands (Sonic Hedgehog (Shh), Desert Hedgehog (Dhh) and Indian Hedgehog (Ihh)) (that all of them are secreted glycoproteins (7)) to membrane receptor called Patched (PTCH). PTCH is a negative regulator of another membrane receptor called smoothened (Smo). During the ligand binding, PTCH inhibition effect on Smo is removed. This event causes alteration of Smo conformation and consequently induction of Gli transcription factors (Gli1, Gli2, and Gli3) will occur. The activity of these transcription factors promotes the transcription of Hh genes, such as Gli1, PTCH, CyclinD1 and Bcl-2 (26, 100-102). The role of Hh signaling pathway (particularly Sonic Hh) in regulating the self-renewal was identified when it was discovered that human HSCs indicate increased self-renewal in response to Sonic Hh stimulation in the laboratory, of course with other growth factors (36, 103). Study of Zhao and coworkers (104) showed that Smo elimination might disturb the ability of HSCs self-renewal and decrease the CML induction by BCR-ABL1 oncoprotein. The Smo elimination was also evacuated in the CML stem cell population, while overexpression of Smo can increase the population of stem cells and facilitate the progression of the disease. The possible mechanism is that inhibitory effects of Smo elimination of LSCs may be related to Numb regulatory effects, which evacuate the CML stem cells.

Notch signaling pathway

Notch signaling pathway plays a role in regulating most of the cellular processes like development and regeneration of mature tissues. Notch membrane receptors are also part of the signaling pathways that are essential for regulating the cell fate of the different tissues; for this reason it is suggested that a disruption in Notch signaling pathway may lead to a dysregulation in the genes that are involved in the self-renewal of the stem cells and finally would cause carcinogenesis and oncogenesis (105). In mammals, Msi function for activating the Notch signaling pathway by suppressing the Numb is reported (106). For the first time, Ito *et al.* pointed that in the CML mouse models, Msi2/Numb expression level and Notch signaling pathway may alter and there is a relationship between them.

They also mentioned that the continued suppression of Numb gene is required to maintain the blast crisis in CML. While, the increasing of Numb expression or the inhibition of Msi2 expression might lead to inhibition of the Notch signaling pathway and direct the cell toward differentiation; therefore, the CML blastic phase is suppressed. The new findings of the Msi2/Numb/Notch signaling pathway may be a key to understanding a new insight to hematopoietic malignancies, especially in the advanced stages of CML blastic crisis (38, 95) (Figure.2 B). Other studies explained the relationship between Notch signaling pathway and the gene for the route (Hes1) with CML displayed progress (107). According to these results, each family member of Musashi is a key regulator of many intracellular signaling pathways and can control the cell fate and stem cell self-renewal, differentiation, and tumorigenesis. Further studies on the relationship between Msi2 and signaling pathways in cancer and various malignancies, including CML is required. Therefore, the expected asymmetric induction of differentiation into mature cells by regulating the signaling pathways involved in self-renewal of cancer cells may provide the new prospect of therapies for leukemia, especially in the advanced stages (37).

Discussion

Inhibition of Msi2 and its ability to reduce the proliferation, cell cycle arrest and induction of apoptosis in leukemia cells

Msi2 gene is proposed as a determinant of cell fate since it induces the rise of cell cycle progression in normal and malignant hematopoietic cells. Zhang *et al.* through MTT assay and Colony formation assays showed that the inhibition of Msi2 reduces duplication and the number of cell clones in the K562 cell line (CML) (73). They reported that Msi2 inhibition leads to an increase of K562 cells in G0/G1 phase and reduction of cells in S phase. The suppression of cell cycle by increasing P21 and decreasing the expression of Cdk2 and Cyclin D1 in both mRNA and protein levels following by inhibiting Msi2 were also demonstrated by them. An increase of P21 expression levels stops cell cycle in G0/G1 phase and causes inhibition of cellular transport from G0/G1 to S phase. In another study, de Andres-Aguayo *et al.* and Hope *et al.* showed that inhibition of Msi2 by increased expression of P21 can significantly reduce mouse HSCs in phase S-G2/M (91, 108). Following the blocking of Msi2 gene, Zhang *et al.* showed early apoptosis induction by flowcytometry staining with Annexin-PI, and the late apoptosis through staining with Wright-Giemsa (for detection of apoptotic cell morphology) (73). To confirm the results of induction of apoptosis, they also reported the reduction in Bcl2 gene expression (an important anti-apoptotic factor) and increase in Bax gene expression (an important factor of apoptosis) in both mRNA and protein levels in K562 cell line. Another study by Kharas *et al.* also showed that Msi2 inhibition by shRNA reduces cell proliferation and significantly induces apoptosis of Nomo-1 and THP-1 (AML), the LAMA-84 and AR230 (CML) cell lines (70).

The importance of targeting the Musashi2-Numb signaling pathway to eradicate leukemia stem cells

CML is a malignant disease having chronic and blast

crisis phases. More recently, appropriate therapies such as kinase inhibitors, Imatinib, were used in clinical base, but this process is not always a successful treatment for patients who are resistant to the medication. As a result, treatment with this drug has some limitations, so new molecular treatment methods are considered (109). Increasing of Msi2 expression levels in advanced stages of CML in comparison with chronic phase of CML patients predicate the increase of unusual activities of the Msi2-Numb signaling pathway; this expanded expression also has been reported in AML. So, inhibition of another purpose apart from BCR-ABL1 (such as Msi2 as an essential gene in the Msi2-Numb) can be considered as a practical approach to eradicate LSCs. This approach by targeting essential genes in signaling pathways that are associated with self-renewal such as Hh and Notch can cause a reduction in aberrant self-renewal of stem cells in CML and prevent the recurrence of the disease (37, 38, 70, 108).

Conclusion

In recent years, considerable progress has been made for treatment of patients with CML. However, providing an approach that is specifically able to reduce LSCs held promise for the effective treatment of CML. The inappropriate activity of Msi2-Numb pathway in LSCs has been reported. Imatinib in the treatment of patients with CML has a significant role, but in spite of this, it has been observed that there are still BCR-ABL1 positive cells that Imatinib is unable to target them; these cancer stem cells (CSCs) are believed to be the cause of resistance to Imatinib therapy. Msi2 is a factor that is expressed at the high levels in CSCs; Msi2 inhibition leads to an increase in the Numb gene expression pattern (as the gene that determines cell fate). This increment of Numb through the key genes of Hh and Notch signaling pathways can decrease the number of LSCs. Therefore, it is obvious that inhibition of Msi2 may be a promising therapeutic approach in reducing human LSCs in CML patients resistant to the drug.

Acknowledgment

We would like to thank all authors responsible for the insights that we attempted to summarize. This work was supported by Tarbiat Modares University.

Conflicts of Interest

The authors declare that there are no conflicts of interest.

References

- Nussbaum RL, McInnes RR, Willard HF. Thompson & Thompson genetics in medicine e-book. Elsevier Health Sciences 2015.
- Sharpless NE, DePinho RA. Cancer biology: gone but not forgotten. *Nature* 2007; 445:606-607.
- Taylor J, Xiao W, Abdel-Wahab O. Diagnosis and classification of hematologic malignancies on the basis of genetics. *Blood* 2017; 130:410-423.
- Toro-Tobón D, Agosto S, Ahmadi S, Koops M, Bruder JM. Chronic myeloid leukemia associated hypercalcemia: a case report and literature review. *Am J Case Rep* 2017; 18:203-207.
- Sultan S, Zaheer HA, Irfan SM, Ashar S. Acute myeloid leukemia: clinical spectrum of 125 patients. *Asian Pac J Cancer Prev* 2016; 17:369-379.
- Keating GM. Dasatinib: a review in chronic myeloid leukaemia and pH+ acute lymphoblastic leukaemia. *Drugs* 2017; 77:85-96.
- Babashah S, Rezaei-Tavirani M, Zamanian-Azodi M, Saki N. Chronic myeloid leukemia as a stem cell-derived malignancy. *J Paramed Sci* 2012; 3:43-55.
- Maru Y. Molecular biology of chronic myeloid leukemia. *Cancer Sci* 2012; 103:1601-1610.
- Sawyers CL. Even better kinase inhibitors for chronic myeloid leukemia. *N Engl J Med* 2010; 362:2314-2315.
- Hantschel O, Superti-Furga G. Regulation of the c-Abl and Bcr-Abl tyrosine kinases. *Nat Rev Mol Cell Biol* 2004; 5:33-44.
- Yuan ZM, Shioya H, Ishiko T, Sun X, Gu J, Huang Y, et al. p73 is regulated by tyrosine kinase c-Abl in the apoptotic response to DNA damage. *Nature* 1999; 399:814-817.
- Druker BJ, Sawyers CL, Kantarjian H, Resta DJ, Reese SF, Ford JM, et al. Activity of a specific inhibitor of the BCR-ABL tyrosine kinase in the blast crisis of chronic myeloid leukemia and acute lymphoblastic leukemia with the Philadelphia chromosome. *N Engl J Med* 2001; 344:1038-1042.
- Calabretta B, Perrotti D. The biology of CML blast crisis. *Blood* 2004; 103:4010-4022.
- Jabbour E, Kantarjian H. Chronic myeloid leukemia: 2016 update on diagnosis, therapy, and monitoring. *Am J Hematol* 2016; 91:252-265.
- Quintás-Cardama A, Cortes JE. Chronic myeloid leukemia: diagnosis and treatment. *Mayo Clin Proc* 2006; 97:81-88.
- Thiele J, Kvasnicka HM, Schmitt-Graeff A, Kriener S, Engels K, Staib P, et al. Effects of the tyrosine kinase inhibitor imatinib mesylate (STI571) on bone marrow features in patients with chronic myelogenous leukemia. *Histol Histopathol* 2004; 19:1277-1288.
- Priyanka A, Mrinal M. The role of new tyrosine kinase inhibitors in chronic myeloid leukemia. *Cancer J* 2016; 22:40-50.
- Walz C, Sattler M. Novel targeted therapies to overcome imatinib mesylate resistance in chronic myeloid leukemia (CML). *Crit Rev Oncol Hematol* 2006; 57:145-164.
- Paul MK, Mukhopadhyay AK. Tyrosine kinase - role and significance in cancer. *Int J Med Sci* 2004; 1:101-115.
- Castagnetti F, Gugliotta G, Breccia M, Iurlo A, Levato L, Albano F, et al. The BCR-ABL1 transcript type influences response and outcome in Philadelphia chromosome-positive chronic myeloid leukemia patients treated frontline with imatinib. *Am J Hematol* 2017; 92:797-805.
- Huang R, Kang Q, Liu H, Li Y. New insights into the molecular resistance mechanisms of chronic myeloid leukemia. *Curr Cancer Drug Targets* 2016; 16:323-345.
- Chen Y, Peng C, Sullivan C, Li D, Li S. Critical molecular pathways in cancer stem cells of chronic myeloid leukemia. *Leukemia* 2010; 24:1545-1554.
- Perrotti D, Silvestri G, Stramucci L, Yu J, Trotta R. Cellular and molecular networks in chronic myeloid leukemia: the leukemic stem, progenitor and stromal cell interplay. *Curr drug targets* 2017; 18:377-388.
- Huntly BJ, Shigematsu H, Deguchi K, Lee BH, Mizuno S, Duclos N, et al. MOZ-TIF2, but not BCR-ABL, confers properties of leukemic stem cells to committed murine hematopoietic progenitors. *Cancer Cell* 2004; 6:587-596.
- Heidel FH, Mar BG, Armstrong SA. Self-renewal related signaling in myeloid leukemia stem cells. *Int J Hematol* 2011; 94:109-117.
- Duman-Scheel M, Weng L, Xin S, Du W. Hedgehog regulates cell growth and proliferation by inducing cyclin D and cyclin E. *Nature* 2002; 417:299-304.
- Boiko AD, Razorenova OV, van de Rijn M, Swetter SM, Johnson DL, Ly DP, et al. Human melanoma-initiating cells

- express neural crest nerve growth factor receptor CD271. *Nature* 2010; 466:133-137.
28. Jamieson CH, Ailles LE, Dylla SJ, Muijtjens M, Jones C, Zehnder JL, *et al.* Granulocyte-macrophage progenitors as candidate leukemic stem cells in blast-crisis CML. *N Engl J Med* 2004; 351:657-667.
 29. Bertacchini J, Ketabchi N, Mediani L, Capitani S, Marmiroli S, Saki N. Inhibition of Ras-mediated signaling pathways in CML stem cells. *Cell Oncol* 2015; 38:407-418.
 30. Catalano A, Rodilossi S, Caprari P, Coppola V, Procopio A. 5-Lipoxygenase regulates senescence-like growth arrest by promoting ROS-dependent p53 activation. *EMBO J* 2005; 24:170-179.
 31. Rådmark O, Werz O, Steinhilber D, Samuelsson B. 5-Lipoxygenase: regulation of expression and enzyme activity. *Trends Biochem Sci* 2007; 32:332-341.
 32. Ghaffari S, Jagani Z, Kitidis C, Lodish HF, Khosravi-Far R. Cytokines and BCR-ABL mediate suppression of TRAIL-induced apoptosis through inhibition of forkhead FOXO3a transcription factor. *Proc Natl Acad Sci U S A* 2003; 100:6523-6528.
 33. Martínez-Gac L, Marqués M, García Z, Campanero MR, Carrera AC. Control of cyclin G2 mRNA expression by forkhead transcription factors: novel mechanism for cell cycle control by phosphoinositide 3-kinase and forkhead. *Mol Cell Biol* 2004; 24:2181-2189.
 34. Miyamoto K, Araki KY, Naka K, Arai F, Takubo K, Yamazaki S, *et al.* Foxo3a is essential for maintenance of the hematopoietic stem cell pool. *Cell Stem Cell* 2007; 1:101-112.
 35. Fasano CA, Dimos JT, Ivanova NB, Lowry N, Lemischka IR, Temple S. shRNA knockdown of Bmi-1 reveals a critical role for p21-Rb pathway in NSC self-renewal during development. *Cell Stem Cell* 2007; 1:87-99.
 36. Reya T, Morrison SJ, MF Clarke, Weissman IL. Stem cells, cancer, and cancer stem cells. *Nature* 2001; 414:105-111.
 37. Nishimoto Y, Okano H. New insight into cancer therapeutics: induction of differentiation by regulating the Musashi/Numb/Notch pathway. *Cell Res* 2010; 20:1083-1085.
 38. Ito T, Kwon HY, Zimdahl B, Congdon KL, Blum J, Lento WE, *et al.* Regulation of myeloid leukaemia by the cell-fate determinant Musashi. *Nature* 2010; 466:765-768.
 39. Kaeda J, Ringel F, Oberender C, Mills K, Quintarelli C, Pane F, *et al.* Up-regulated MSI2 is associated with more aggressive chronic myeloid leukemia. *Leuk Lymphoma* 2015; 56:2105-2113.
 40. Bumbea H, Vladareanu AM, Voican I, Cisleanu D, Barsan L, Onisai M. Chronic myeloid leukemia therapy in the era of tyrosine kinase inhibitors-the first molecular targeted treatment. *J Med Life* 2010; 15:162-166.
 41. Pavlu J, Szydlo RM, Goldman JM, Apperley JF. Three decades of transplantation for chronic myeloid leukemia: what have we learned?. *Blood* 2011; 117:755-763.
 42. Sullivan C, Peng C, Chen Y, Li D, Li S. Targeted therapy of chronic myeloid leukemia. *Biochem pharmacol* 2010; 80:584-591.
 43. Talpaz M, Hehlmann R, Quintana-Cardama A, Mercer J, Cortes J. Re-emergence of interferon- α , in the treatment of chronic myeloid leukemia. *Leukemia* 2013; 27:803-812.
 44. Wetzel R, Goss VL, Norris B, Popova L, Melnick M, Smith BL. Evaluation of CML model cell lines and imatinib mesylate response: determinants of signaling profiles. *J Immunol Methods* 2005; 305:59-66.
 45. Kayastha GK, Ranjitkar N, Gurung R, KC R, Karki S, Shrestha R, *et al.* The use of Imatinib resistance mutation analysis to direct therapy in Philadelphia chromosome/BCR-ABL1 positive chronic myeloid leukaemia patients failing Imatinib treatment, in Patan hospital, Nepal. *Br J Haematol* 2017; 177:1000-1007.
 46. Bixby D, Talpaz M. Seeking the causes and solutions to imatinib-resistance in chronic myeloid leukemia. *Leukemia* 2011; 25:7-22.
 47. Rousselot P, Huguet F, Rea D, Legros L, Cayuela JM, Maarek O, *et al.* Imatinib mesylate discontinuation in patients with chronic myelogenous leukemia in complete molecular remission for more than 2 years. *Blood* 2007; 109:58-60.
 48. Dierks C, Beigi R, Guo GR, Zirlik K, Stegert MR, Manley P, *et al.* Expansion of Bcr-Abl-positive leukemic stem cells is dependent on Hedgehog pathway activation. *Cancer cell* 2008; 14:238-249.
 49. Mahon FX, Réa D, Guilhot J, Guilhot F, Huguet F, Nicolini F, *et al.* Discontinuation of imatinib in patients with chronic myeloid leukaemia who have maintained complete molecular remission for at least 2 years: the prospective, multicentre Stop Imatinib (STIM) trial. *Lancet Oncol* 2010; 11:1029-1035.
 50. Cortes J, Hochhaus A, Hughes T, Kantarjian H. Front-line and salvage therapies with tyrosine kinase inhibitors and other treatments in chronic myeloid leukemia. *J Clin Oncol* 2011; 29:524-531.
 51. Verga Falzacappa MV, Ronchini C, Reavie LB, Pelicci PG. Regulation of self-renewal in normal and cancer stem cells. *FEBS J* 2012; 279:3559-3572.
 52. Dick D. Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. *Nature Med* 1997; 3:730-737.
 53. Chu S, Xu H, Shah NP, Snyder DS, Forman SJ, Sawyers CL, *et al.* Detection of BCR-ABL kinase mutations in CD34+ cells from chronic myelogenous leukemia patients in complete cytogenetic remission on imatinib mesylate treatment. *Blood* 2005; 105:2093-2098.
 54. Graham SM, Vass JK, Holyoake TL, Graham GJ. Transcriptional analysis of quiescent and proliferating CD34+ human hemopoietic cells from normal and chronic myeloid leukemia sources. *Stem Cells* 2007; 25:3111-3120.
 55. Graham SM, Jørgensen HG, Allan E, Pearson C, Alcorn MJ, Richmond L, *et al.* Primitive, quiescent, Philadelphia-positive stem cells from patients with chronic myeloid leukemia are insensitive to STI571 *in vitro*. *Blood* 2002; 99:319-325.
 56. Jørgensen HG, Allan EK, Jordanides NE, Mountford JC, Holyoake TL. Nilotinib exerts equipotent antiproliferative effects to imatinib and does not induce apoptosis in CD34+ CML cells. *Blood* 2007; 109:4016-4019.
 57. Houghton J, Morozov A, Smirnova I, Wang TC. Stem cells and cancers. *Seminars in cancer biology*. Elsevier 2007; 19:1-203.
 58. Horne GA, Jackson L, Helgason V, Holyoake TL. Stem cell guardians—old and new perspectives in LSC biology. *Curr Drug Targets* 2017; 18:405-413.
 59. Naka K, Hoshii T, Hirao A. Novel therapeutic approach to eradicate tyrosine kinase inhibitor resistant chronic myeloid leukemia stem cells. *Cancer Sci* 2010; 101:1577-1581.
 60. Park CY, Tseng D, Weissman IL. Cancer stem cell-directed therapies: recent data from the laboratory and clinic. *Mol Ther* 2009; 17:219-230.
 61. Huntly BJ, Gilliland DG. Leukemia stem cells and the evolution of cancer-stem-cell research. *Nat Rev Cancer* 2005; 5:311-321.
 62. Krause DS, Van Etten RA. Right on target: eradicating leukemic stem cells. *Trends Mol Med* 2007; 13:470-481.
 63. Jamieson CH. Chronic myeloid leukemia stem cells. *ASH Education Program Book* 2008; 436-442.
 64. Murat A, Migliaiaccia E, Gorlia T, Lambiv WL, Shay T, Hamou MF, *et al.* Stem cell-related "self-renewal" signature and high epidermal growth factor receptor expression associated with resistance to concomitant chemoradiotherapy in glioblastoma. *J Clin Oncol* 2008; 26:3015-3024.
 65. Al-Hajj M, Clarke MF. Self-renewal and solid tumor stem cells. *Oncogene* 2004; 23:7274-7282.
 66. Lobo NA, Shimono Y, Qian D, Clarke MF. The biology of

- cancer stem cells. *Annu Rev Cell Dev Biol* 2007; 23:675-699.
67. Tan BT, Park CY, Ailles LE, Weissman IL. The cancer stem cell hypothesis: a work in progress. *Lab Invest* 2006; 86:1203-1207.
68. Zhang H, Li S. Molecular mechanisms for survival regulation of chronic myeloid leukemia stem cells. *Protein Cell* 2013; 4:186-196.
69. Chavez-Gonzalez A, Bakhshinejad B, Pakravan K, Guzman ML, Babashah S. Novel strategies for targeting leukemia stem cells: sounding the death knell for blood cancer. *Cell Oncol* 2017; 40:1-20.
70. Kharas MG, Lengner CJ, Al-Shahrour F, Bullinger L, Ball B, Zaidi S, *et al.* Musashi-2 regulates normal hematopoiesis and promotes aggressive myeloid leukemia. *Nature med* 2010; 16:903-908.
71. Sakakibara SI, Nakamura Y, Satoh H, Okano H. RNA-binding protein Musashi2: developmentally regulated expression in neural precursor cells and subpopulations of neurons in mammalian CNS. *J Neurosci* 2001; 21:8091-8107.
72. RG de Bruin. TJ Rabelink. AJ van Zonneveld. EP van der Veer. Emerging roles for RNA-binding proteins as effectors and regulators. *Eur Heart J* 2017; 38:1380-1388.
73. Zhang H, Tan S, Wang J, Chen S, Quan J, Xian J, *et al.* Musashi2 modulates K562 leukemic cell proliferation and apoptosis involving the MAPK pathway. *Exp Cell Res* 2014; 320:119-127.
74. Okano H, Kawahara H, Toriya M, Nakao K, Shibata S, Imai T. Function of RNA-binding protein Musashi-1 in stem cells. *Exp Cell Res* 2005; 306:349-356.
75. Sugiyama-Nakagiri Y, Akiyama M, Shibata S, Okano H, Shimizu H. Expression of RNA-binding protein Musashi in hair follicle development and hair cycle progression. *Am J Pathol* 2006; 168:80-92.
76. Sakakibara S, Nakamura Y, Yoshida T, Shibata S, Koike M, Takano H, *et al.* RNA-binding protein Musashi family: roles for CNS stem cells and a subpopulation of ependymal cells revealed by targeted disruption and antisense ablation. *Proc Natl Acad Sci USA* 2002; 99:15194-15199.
77. Sakakibara S, Imai T, Hamaguchi K, Okabe M, Aruga J, Nakajima K, *et al.* Mouse-Musashi-1, a neural RNA-binding protein highly enriched in the mammalian CNS stem cell. *Dev Biol* 1996; 176:230-242.
78. Sakakibara S, Okano H. Expression of neural RNA-binding proteins in the postnatal CNS: implications of their roles in neuronal and glial cell development. *J Neurosci* 1997; 17:8300-8312.
79. Akasaka Y, Saikawa Y, Fujita K, Kubota T, Ishikawa Y, Fujimoto A, *et al.* Expression of a candidate marker for progenitor cells, Musashi-1, in the proliferative regions of human antrum and its decreased expression in intestinal metaplasia. *Histopathology* 2005; 47:348-356.
80. Kayahara T, Sawada M, Takaishi S, Fukui H, Seno H, Fukuzawa H, *et al.* Candidate markers for stem and early progenitor cells, Musashi-1 and Hes1, are expressed in crypt base columnar cells of mouse small intestine. *FEBS Lett* 2003; 535:131-135.
81. Potten CS, Booth C, Tudor GL, Booth D, Brady G, Hurley P, *et al.* Identification of a putative intestinal stem cell and early lineage marker; musashi-1. *Differentiation* 2003; 71:28-41.
82. Clarke RB, Spence K, Anderson E, Howell A, Okano H, Potten CS. A putative human breast stem cell population is enriched for steroid receptor-positive cells. *Dev Biol* 2005; 277:443-456.
83. Li D, Peng X, Yan D, Tang H, Huang F, Yang Y, *et al.* Msi-1 is a predictor of survival and a novel therapeutic target in colon cancer. *Ann Surg Oncol* 2011; 18:2074-2083.
84. Wang XY, Penalva LO, Yuan H, Linnoila RI, Lu J, Okano H, *et al.* Musashi1 regulates breast tumor cell proliferation and is a prognostic indicator of poor survival. *Mol Cancer* 2010; 22:13-12.
85. Nakano A, Kanemura Y, Mori K, Kodama E, Yamamoto A, Sakamoto H, *et al.* Expression of the neural RNA-binding protein Musashi1 in pediatric brain tumors. *Pediatr Neurosurg* 2007; 43:279-284.
86. Nikpour P, Baygi ME, Steinhoff C, Hader C, Luca AC, Mowla SJ, *et al.* The RNA binding protein Musashi1 regulates apoptosis, gene expression and stress granule formation in urothelial carcinoma cells. *J Cell Mol Med* 2011; 15:1210-1224.
87. Kanemura Y, Yamasaki M, Mori K, Fujikawa H, Hayashi H, Nakano A, *et al.* Musashi1, an evolutionarily conserved neural RNA-binding protein, is a versatile marker of human glioma cells in determining their cellular origin, malignancy, and proliferative activity. *Differentiation* 2001; 68:141-152.
88. Bobryshev Y, Freeman A, Botelho N, Tran D, Levert-Mignon A, Lord R. Expression of the putative stem cell marker Musashi-1 in Barrett's esophagus and esophageal adenocarcinoma. *Dis Esophagus* 2010; 23:580-589.
89. Ye F, Zhou C, Cheng Q, Shen J, Chen H. Stem-cell-abundant proteins Nanog, Nucleostemin and Musashi1 are highly expressed in malignant cervical epithelial cells. *BMC Cancer* 2008; 8:108.
90. Toda M, Iizuka Y, Yu W, Imai T, Ikeda E, Yoshida K, *et al.* Expression of the neural RNA-binding protein Musashi1 in human gliomas. *Glia* 2001; 34:1-7.
91. Hope KJ, Cellot S, Ting SB, MacRae T, Mayotte N, Iscove NN, *et al.* An RNAi screen identifies Msi2 and Prox1 as having opposite roles in the regulation of hematopoietic stem cell activity. *Cell Stem Cell* 2010; 7:101-113.
92. Sureban SM, May R, George RJ, Dieckgraefe BK, McLeod HL, Ramalingam S, *et al.* Knockdown of RNA binding protein musashi-1 leads to tumor regression *in vivo*. *Gastroenterology* 2008; 134:1448-1458.
93. Moreira AL, Gonen M, Rekhtman N, Downey RJ. Progenitor stem cell marker expression by pulmonary carcinomas. *Mod Pathol* 2010; 23:889-895.
94. de Andrés-Aguayo L, Varas F, Graf T. Musashi 2 in hematopoiesis. *Curr Opin hematol* 2012; 19:268-272.
95. Moore MA. A cancer fate in the hands of a samurai. *Nature medicine* 2010; 16:963-965.
96. MacNicol AM, Wilczynska A, MacNicol MC. Function and regulation of the mammalian Musashi mRNA translational regulator. *Biochem Soc Trans* 2008; 36:528-530.
97. Cohen MM. The hedgehog signaling network. *Am Journal Med Genet A* 2003; 123:5-28.
98. Lewis MT, Veltmaat JM. Next stop, the twilight zone: hedgehog network regulation of mammary gland development. *J Mammary Gland Biol Neoplasia* 2004; 9:165-181.
99. Ingham PW, McMahon AP. Hedgehog signaling in animal development: paradigms and principles. *Genes Dev* 2001; 15:3059-3087.
100. Yang L, Xie G, Fan Q, Xie J. Activation of the hedgehog-signaling pathway in human cancer and the clinical implications. *Oncogene* 2010; 29:469-481.
101. Varjosalo M, Taipale J. Hedgehog signaling. *J Cell Sci* 2007; 120:3-6.
102. Nehmé R, Mus-Veteau I. Proteins of the Hedgehog signaling pathway as therapeutic targets against cancer. *Expert Rev Proteomics* 2010; 7:601-612.
103. Babashah S, Sadeghizadeh M, Hajifathali A, Tavirani MR, Zomorod MS, Ghadiani M, *et al.* Targeting of the signal transducer Smo links microRNA-326 to the oncogenic Hedgehog pathway in CD34+ CML stem/progenitor cells. *Int J Cancer* 2013; 133:579-589.
104. Zhao C, Chen A, Jamieson CH, Fereshteh M, Abrahamsson A, Blum J, *et al.* Hedgehog signalling is essential for

maintenance of cancer stem cells in myeloid leukemia. *Nature* 2009; 458:776-779.

105. Hambardzumyan D, Becher OJ, Holland EC. Cancer stem cells and survival pathways. *Cell Cycle* 2008; 7:1371-1378.

106. Imai T, Tokunaga A, Yoshida T, Hashimoto M, Mikoshiba K, Weinmaster G, *et al.* The neural RNA-binding protein Musashi1 translationally regulates mammalian numb gene expression by interacting with its mRNA. *Mol Cell Biol* 2001; 21:3888-3900.

107. Nakahara F, Sakata-Yanagimoto M, Komeno Y, Kato N,

Uchida T, Haraguchi K, *et al.* Hes1 immortalizes committed progenitors and plays a role in blast crisis transition in chronic myelogenous leukemia. *Blood* 2010; 115:2872-2881.

108. Byers RJ, Currie T, Tholouli E, Rodig SJ, Kutok JL. MSI2 protein expression predicts unfavorable outcome in acute myeloid leukemia. *Blood* 2011; 118:2857-2867.

109. Scherr M, Battmer K, Blömer U, Schiedlmeier B, Ganser A, Grez M, *et al.* Lentiviral gene transfer into peripheral blood-derived CD34+ NOD/SCID-repopulating cells. *Blood* 2002; 99:709-712.